

Integrated use of the next generation plant biostimulants for an enhanced sustainability of field vegetable high residue farming systems –STIM 4⁺ RO-NO-2019-540

D. Constantinescu-Aruxandei, S. Brooks, A. Nicolescu, S. Shaposhnikov, F. Georgescu, L.A. Pairault, L. Marin, C. Deleanu, F. Oancea





Partners

- National Institute for Research & Development in Chemistry and Petrochemistry—ICECHIM, Bucharest, Romania
- Norway Institute for Water Research—NIVA, Oslo, Norway
- Petru Poni Institute for Macromolecular Chemistry, Iasi, Romania
- Norgenotech AS, Skreia, Norway
- Enpro Soctech, Bucharest, Romania
- Amia Import-Export, Otopeni, Romania



Overall aims and objectives of the project.

The goal of the STIM4⁺ project is to develop (bio)technologies for the production and the integrated utilization of next generation plant biostimulants, for field vegetables grown into high residue farming systems.

The STIM 4⁺ project objectives are:

(*i*) To develop next generation plant biostimulants intended to improve the resource use efficiency of high-residue grown vegetables;

(*ii*) To assess and to characterize the new plant biostimulants effects on vegetables and rhizosphere microorganism;

(*iii*) To investigate the safety and environmental impact of the next generation plant biostimulants.



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> Trade-off of the vegetable cultivation into high residues system - winter cover crop mulch

Negative impacts ↑ Soil borne crop diseases
 ↑ Weed (including parasitic one) infestation

↓Nutrients (mainly nitrogen) availability

↓Heavy soil structure

↓ Soil temperature (temperate climate)

Crop Production

↑ Water storage ↑ Water use efficiency

↑ Light soil structure

↑ Organic matter

↑Storage of nutrients
↑ Biological diversity

Main advantages

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Next-generation plant biostimulants

(*i*) multi-functional *Trichoderma* strains-based plant biostimulants (a microbial plant biostimulants);

(*ii*) glycodynameric, chitosan based bioactive (micro)hydrogel formulation (organic plant biostimulants);

(*iii*) zerovalent selenium nanoparticles (inorganic plant biostimulant).(*iv*) microalgae standardized extract, including phytohormones, polyamines and betaines (organic plant biostimulant), fortified with a strigolactone mimic (bio-designed plant biostimulant);





Next-generation plant biostimulants to compensate high residues farming drawbacks







Voges–Proskauer test

IAA production



Nitrilase production

Phosphorus

Phosphorus solubilization

in vitro production of plant biostimulant compounds

Siderophores

production

cytokinin production



ACC-deaminase

Polyamines production

Secretion of glycosyl hydrolases and solubilization of phyto-silica



Production of polypeptides which are amplifying the cellulases activities



Selected microbial strains



Testing multiple interactions



Selection of the multifunctional consortia

Selection of multi-functional *Trichoderma* strains

HTS to select *Trichoderma* consortium responding to strigolactone



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	0	T50	Т36	T27	Td1	Si16	Tk20	Mt	Si7	Tk14	T83	102
	T57	Tk14	Т83	Т36	Td2	T50	Si16	Т27	Td1	Si7	Tk20	Mt-
	Mt-	Si7	Tk20	т83	T57	Tk14	т50	Т36	Td2	Td1	Si16	T27
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Consortium Si7- T27 - Tk20

Patent RO131827 - PCT application

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Glycodynameric, chitosan based bioactive (micro)hydrogel formulation

Iftime, M. M., Rosca, I., Sandu, A. I., & Marin, L. (2022). Chitosan crosslinking with a vanillin isomer toward self-healing hydrogels with antifungal activity. *International Journal of Biological Macromolecules*, *205*, 574-586. SOIL-

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Preparation of SL-20 -loaded chitosan-salicyl-imine hydrogels



Degradation in soil of the formulations and swelling capacity

Code	Time (days)	1 day	7 days	2 days	7/9 days			
Со	8			-	*	MES	xerogels	
						Probe	1 di	av
C1					·	Со	12	-,
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C3		14.19	9 days			CS-SL	29	
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C5	15							
CS- SI20	4			6				

MES	xerogels	pellets	Degradation time (day		
Probe	1 da	ау	xerogels	pellets	
Со	12	8	8	8	
C1	16	10	8	8	
C2	14	12	9	10	
C3	13	11	9	10	
C4	8	7	10	12	
C5	5	4	15	22	
CS-SL	29	-	4	1	

mages of the formulations sample in soil, before and after degradation experiment

Glycodynameric, chitosan based bioactive (micro)hydrogel formulation

In order to choose the glycodinameric hydrogel for further developing of the biostimulant formulation, a series of properties, relevant for formulation obtaining and behaviour, were investigated. They are briefly presented below.

Hydrogel morphology influences their ability to retain water in soil, to encapsulate different bioactive compounds and to release them in a controlled manner. The morphology of the hydrogels was assessed by scanning electron microscopy. It was observed that they have a porous structure, with well delimited pores for the hydrogels with high crosslinking density, and a fibrous structure for those with lower crosslinking density.



Representative SEM images of hydrogels at different magnifications a) 1000x and b) 150x

Novel, identified microbial strains which produce nanoselenium



✓ Starting from commercially available 1,8-naphthalic anhydrides:



√ Starting from 4-(4-hydroxyphenyl)pyrimidines:



8 (SL-13)



Solvatochromic effect

- Nicolescu, A., Airinei, A., Georgescu, E., Georgescu, F., Tigoianu, R., Oancea, F., & Deleanu, C. (2021). Synthesis, photophysical properties and solvatochromic analysis of some naphthalene-1, 8-dicarboxylic acid derivatives. *Journal of Molecular Liquids, 303*, 112626.
- Tigoianu, R., Airinei, A., Georgescu, E., Nicolescu, A., Georgescu, F., Isac, D. L., Deleanu, C. & Oancea, F. (2022). Synthesis and solvent dependent fluorescence of some piperidinesubstituted naphthalimide derivatives and consequences for water sensing. *International journal of molecular sciences*, 23(5), 2760.

Advanced structural characterization of the newly synthesized strigolactone mimics

Types of NMR experiments performed during activity T2.1, exemplified for compound SL-20. All the experiments were recorded on a Bruker Avance NEO 600MHz spectrometer.





Acute toxicity on earthworm - ISO 11268-1:2012



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Ecotoxicity evaluation of Strigolactone (SL-6)

- Freshwater toxicity:
 - Unicellular algae growth inhibitio (OECD201)
 - Daphnia magna (OECD202)
 - Fish embryo test (OECD236)



- Marine toxicity:
 - Algal growth inhibition (ISO10253)
 - Fucus germling growth
 - Copepod (Tisbe) acute toxicity (ISO14699)
 - Oyster embryo bioassay (ASTM E724-89)







Genotoxicity of SL-6 in Microalgae



model



Cell genotoxicity (% tail DNA) of SL-6 to the microalgae model. C-control, SC-solvent control (SL-6 concentration in mg/L, mean \pm SD). Method 1: Standard lysis, 2.5 M NaCl, 0.1 M Na₂EDTA, 10 mM Tris Base, 1% Triton-X-100, pH 10. Method 2: Alternative lysis, 300 mM NaOH, 2 mM Na2 EDTA, 0.01% SDS. Subsequent steps were as for the standard comet assay for strand breaks. Statistically significant increase (*p<0.05, **p<0.01, ***p<0.001)



Presentation of the integrated process to produce nanoselenium and microalgae based plant biostimulants. The microalgae bacteria are grown in the presence of strigolactone and 100 μ g/L Na₂SeO₃ for 14 days. The biomass is harvested, SeNPs are separated from the cell components and cell components are used as an ingredient of the organic plant biostimulant, fortified with a strigolactone mimic.

Field experiment



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Increased yield of field grown tomatoes С

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Improved fruit quality





Easy extractable glomalin and soil nitrate







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Polyamines and strigolactones -Stimulation of mycorrhizal symbiosis formation

Constantinescu-Aruxandei, D., & Oancea, F. (2023). Closing the Nutrient Loop—The New Approaches to Recovering Biomass Minerals during the Biorefinery Processes. *International Journal of Environmental Research and Public Health, 20*(3), 2096.

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